



A Systematic Review Exploring the **Anticancer Activity and Mechanisms** of Glucomannan

Jun-yi Li^{1†}, Fei Sun^{2†}, Hai-feng Zhou^{2†}, Jia Yang¹, Cong Huang³ and Heng Fan^{1*}

¹ Department of Integrated Traditional Chinese and Western Medicine, Union Hospital, Tongji Medical College, Huazhong University of Science and Technology, Wuhan, China, ² The Center for Biomedical Research, Tongji Hospital, Tongji Medical College, Huazhong University of Science and Technology, Wuhan, China, ³ Institute of Integrated Traditional Chinese and Western Medicine, Tongji Hospital, Tongji Medical College, Huazhong University of Science and Technology, Wuhan, China

Glucomannan, long recognized as the active ingredient of the traditional Chinese medicinal herb Konjac glucomannan, is a naturally occurring polysaccharide existing in certain plant species and fungi. Due to its special property to also serve as a dietary supplement, glucomannan has been widely applied in clinic to lower body weight and circulation cholesterol level and to treat constipation, diabetes, and arterial sclerosis. Besides the regulatory role engaged with gastroenterological and metabolic syndrome, recently, its therapeutic effect and the underlying mechanisms in treating cancerous diseases have been appreciated by mounting researches. The present review aims to emphasize the multifaceted aspects of how glucomannan exerts its anti-tumor function.

OPEN ACCESS

Edited by:

Jiangjiang Qin, Zhejiang Chinese Medical University, China

Reviewed by:

Qi Zena. Xidian University, China Qiang Wang. Nanjing Drum Tower Hospital, China

*Correspondence:

Heng Fan fanheng009@aliyun.com

[†]These authors contributed equally to this work.

Specialty section:

This article was submitted to Pharmacology of Anti-Cancer Drugs, a section of the journal Frontiers in Pharmacology

> Received: 11 June 2019 Accepted: 22 July 2019 Published: 23 August 2019

Citation:

Li J-y, Sun F, Zhou H-f, Yang J, Huang C and Fan H (2019) A Systematic Review Exploring the Anticancer Activity and Mechanisms of Glucomannan. Front. Pharmacol. 10:930. doi: 10.3389/fphar.2019.00930

Keywords: glucomannan, polysaccharide, apoptosis, drug carrier, anti-tumor effect

INTRODUCTION

Currently, tumors are increasingly becoming a global health issue. According to an epidemiological survey, cancer already surpasses cardiovascular and cerebrovascular accidents, and ranks as the top life-threatening disease worldwide. A status report on the global burden of cancer using GLOBOCAN estimated 18.1 million new cancer cases and 9.6 million cancer deaths in 2018 year alone (Bray et al., 2018). The overall cancer (containing more than 18 types of cancer) 5-year survival rates in China, Japan, and the United States were 36.0%, 57.4%, and 64.0% (Allemani et al., 2018), which suggests a significant variation among different countries.

The reasons that make cancer so tough to deal with are complicated. Basically speaking, dysfunctional cell cycle regulation with an excessive proliferation capacity and undifferentiated immature phenotype are the hallmarks of cancer pathology. Besides, tumor cells show intrinsic aberrance of multiple cellular processes. High expression of pro-survival genes like BCL-2 and MCL1 render them apoptosis resistant (Sawai et al., 2018), and metabolic pathways are rewired to make tumor cells better adapt to the changing environment. Tumor cells are experts in nutrient utilization; they adopt distinct metabolic programs, ranging from glycolysis, fatty acid oxidation, to amino acid metabolism under various circumstances. Accompanied by the elevated metabolic activity required for uncontrolled growth, the production of reactive oxygen species (ROS) is substantially increased, which further causes genomic instability and accumulation of genetic mutation (Sabharwal and Schumacker, 2014). Interestingly, autophagy seems to play a dual role in tumor development. During the early stage, autophagy inhibits tumor formation through alleviating cellular stress; however, this mechanism is then hijacked by later-stage tumor cells to facilitate their growth under extreme conditions. To make things awkward, tumors could also have active impact

on the microenvironment where they live. VEGF-C from tumor cells promotes blood and lymphatic vessel neogenesis, which is in favor of tumor cells' development and metastasis (Liu et al., 2019). Meantime, immune cells are educated into a tolerant or anergic state, thus leading to tumor immune evasion.

Researches on cancer treatment are focused on two aspects: one is to discover new anti-tumor therapy, while another is to establish an efficient drug-delivery system. With the advance of medical science, we now possess huge artillery of weapons against tumor. Conventional protocols including surgical removal, radiotherapy, and chemotherapy form the basis of clinical therapies. Strikingly, immunological approaches are emerging as a strong supplement. The deployment of cell-based CAR-T (Abramson et al., 2017) or DC-CIK strategy (Zhang et al., 2018) and usage of checkpoint inhibitors (anti-PD-1, anti-CTLA4, etc.) (Wei et al., 2017) as well as tumor vaccines (Kaiser, 2017) bring about great benefits for certain populations of patients. To further improve the selectivity, biomarker-discovery-based monoclonal antibody development and finely designed nanoparticle carriers are applied. Nonetheless, these measures are not enough to win the battle against tumors. The side effect, off-target effect, and loss of effect are prevalent phenomena among present therapeutic protocols. Thus, it is of importance to pursue additional methods and bioactive anti-tumor compounds.

Glucomannan is a family of polysaccharide widely existing in higher plants and microorganisms. The backbone structure is β -(1>4) glycoside bond linked D-mannose and D-glucose in a ratio of 1.6:1, while it is only lightly branched through β -(1 \rightarrow 6) glucosyl moieties (Wu et al., 2011a). The molecular weight ranges from 200,000 to 2,000,000 Daltons, which varies with origin, method of processing, and storage time. In general, glucomannan has favorable characteristics of water solubility and extremely low toxicity (Hassan et al., 2014; Luan et al., 2017), which make it a fantastic bio-compatible compound. Actually, Konjac glucomannan, the well-known member of the glucomannan family, was first documented as a traditional Chinese herb about 2000 years ago (CP C, 2010). Since then, it has been used to treat conditions such as asthma, cough, hernia, breast pain, burns, and hematological and skin diseases (CP C, 2010). Nowadays, it mainly serves either as a dietary supplement aiding in weight loss, diabetes, arterial sclerosis, and constipation, or as an emulsifier and thickener in food processing (Chen et al., 2017a; Wu et al., 2018), which corroborates the old theory of "homology of medicine and food." In fact, many natural polysaccharides that show potent anti-tumor activity (Kim et al., 2011; Xiao et al., 2017; Zhang, 2017; Deng et al., 2018) derive from edible herbs, such as Lentinus, Cordyceps, Ganoderma lucidum, and Hericium, which themselves are also common cuisine ingredients. Recently, glucomannan extracted from Amorphophallus konjac (Chen et al., 2017a), Bletilla striata (Zhan et al., 2014), Lentinus edodes (Fujii et al., 1978), Aloe vera (Sampedro et al., 2004; Im et al., 2005; Liu et al., 2006; Im et al., 2016; Quezada et al., 2017), and Candida utilis (Kumano et al., 1985) were demonstrated to have similar effect. Moreover, according to the studies reviewed in the present research, glucomannan, mainly from these species, not only is an anti-cancer drug by itself but also works as a targeted carrier that is compatible with various bioactive compounds.

GLUCOMANNAN DIRECTLY INTERFERES WITH INTRINSIC TUMOR CELLS BIOLOGICAL PROCESSES

Glucomannan Impairs Tumor Cells' Survival and Metastasis

Given that tumor cells are apoptosis resistant, drugs aimed to promote apoptosis are widely used in cancer therapy (Hassan et al., 2014). Initially, studies unveiled the therapeutic effect of extract from A. konjac (AKe) on various cancers, such as colon carcinoma (Ansil et al., 2013; Ansil et al., 2014a), gastric cancer (Chen et al., 2017a), hepatoma (Ansil et al., 2014b), and breast cancer (Wu et al., 2018; Wu et al., 2019). Chen et al. found that AKe could inhibit the growth of in vitro cultured gastric cancer cell lines SGC-7901 and AGS (Chen et al., 2017a). On the molecular level, the expression of inhibitors of apoptosis protein (IAP) family member survivin and the pro-survival gene BCL2 decreased, while the pro-apoptosis protein BAX and caspase-9 increased (Chen et al., 2017a) (Figure 1). AKe also displayed similar therapeutic effect on human liver cancer cell line and the human triple-negative breast cancer (TNBC) cells (Ansil et al., 2014b; Wu et al., 2018). By blocking the transition from G0/G1 to G2/M phase and decreasing the expression of the proliferative marker proliferative cell nuclear antigen (PCNA), AKe promotes cell cycle arrest and inhibits cell division (Ansil et al., 2013; Chen et al., 2017a) (Figure 1). The elevated activity of tumor cell apoptosis is always accompanied by decreased cell proliferation, which stems from the shared down-regulated PI3K/AKT signaling pathway (Frisch and Screaton, 2001; Guanen et al., 2018). Since Konjac glucomannan was identified as the major bioactive component of the A. konjac (Chen et al., 2017a), not surprisingly, Sawai S. and his colleagues found that Konjacglucomannan-treated HepG2 hepatic carcinoma cells displayed a significant reduction of growth, which resembles the phenotype of AKe-treated tumor cells (Sawai et al., 2018).

Indeed, the PI3K/AKT/mTOR pathway is frequently hyperactivated in tumor cells and also plays an important role in cancer metastasis (Guanen et al., 2018). AKe was found to inhibit PI3K signaling pathway and thwart MDA-MB231 breast cancer cell metastasis to lung in a xenograft model (Wu et al., 2018). The chemokine receptors and adhesion molecules are strongly linked to tumor recurrence and metastasis. AKe was reported to significantly increase E-cadherin expression and reduce CCR7 and CXCR4 level, which are required for tumor migration (Wu et al., 2018) (**Figure 1**). Moreover, the glucomannan extracted from the *C. utilis* preferentially inhibited the Lewis lung carcinoma (3LL) pulmonary metastases in a time-dependent manner, wherein an optimal dosage of glucomannan on early stage exhibits the optimal suppressive effect (Kumano et al., 1985).

Collectively, these data outline the intrinsic impacts of glucomannan on tumor cell apoptosis, proliferation, and metastatic activity, which are the essential basis for its antitumor function. To a large extent, such effects are attributed to glucomannan-mediated inhibition of the PI3K/AKT signaling pathway. As a result, increased ratio of pro-apoptotic to



anti-apoptotic gene expression undermines the immortality of tumor cells, while the reduced level of CCR7 and CXCR4 compromises their migratory capability.

Glucomannan Enhances Anti-Oxidative Activity and Induces Autophagy in Tumor Cells

Another glucomannan-mediated anti-cancer effect could be ascribed to its role as an antioxidant inducer (Miadoková et al., 2006; Ansil et al., 2013; Ansil et al., 2014b; Wu et al., 2014). An earlier study confirmed that glucomannan isolated from C. utilis exerts anti-mutagenic function in a mouse model of cyclophosphamideinduced mutagenicity, which may result from the decreased ROS level (Chorvatovicova et al., 1999). Miadoková et al. further concluded that glucomannan from C. utilis cell wall exerts the anti-oxidative activity through iron chelation and scavenging of hydroxyl radicals in mouse leukemia cell (Miadoková et al., 2006). In line with the above observation, glucomannan or AKe was demonstrated to enhance the expression of antioxidant enzymes in 1,2-dimethylhydrazine (DMH)-induced colon carcinogenesis and N-nitrosodiethylamine-induced hepatocellular carcinoma in rats (Ansil et al., 2013; Ansil et al., 2014c). Besides, Konjac glucomannan was able to ameliorate AOM-induced genotoxicity via alleviating the DNA damage process initiated by accumulated ROS (Wu et al., 2014) (Figure 1).

Alternatively, in the gastric cancer cell lines SGC-7901 and AGS, investigators found that AKe treatment markedly increased the expression of LC3-II, which is a marker of the mature autophagosome and indicates high level of autophagy

(Chen et al., 2017a). Increased cell autophagy plays a protective role in early tumor development, with the possible mechanisms of autophagy-related stress elimination and even autophagyinduced cell death. Nassour et al. found that autophagy is an important cellular protective process to induce cell death during replicative crisis and impaired autophagy is required for initiation of tumorigenesis (Nassour et al., 2019). Though the AKT/mTOR pathway may be related to the activation of the autophagic process (Chen et al., 2014), there is no definite evidence showing the existence of the glucomannan-AKT/mTOR-autophagy axis in the tumor cells.

GLUCOMANNAN INDIRECTLY AFFECTS TUMOR DEVELOPMENT

Glucomannan Promotes Extrinsic Environment Disfavoring Tumorigenesis

Glucomannan displays a beneficial effect for constipation and promotes intestinal peristalsis. As a dietary water-absorbing fiber, it has also been demonstrated to reduce the production of carcinogens from the gut (Mizutani and Mitsuoka, 1982; Wu and Chen, 2011b; Wu et al., 2011a; Chong, 2013). Due to its hydrolytic ability of turning substances into carcinogens, β -glucuronidase is identified as a lysosomal enzyme involved in the process of tumor development and metastasis (Klinder et al., 2008). Another protein peptide hydrolase, mucinase, is able to hydrolyze the mucin layer of the gut lumen and make the enterocytes to be in direct contact with colon carcinogens

(Miller and Hoskins, 1981). According to Dr. Wu's research, the incorporation of Konjac glucomannan in high-fat fiber-free diet in rats decreased the activity of β -glucuronidase and mucinase by 71% and 68%, respectively (Wu and Chen, 2011b) (**Figure 2**).

Studies proved that bile acid is in close relationship with the carcinogenesis process (Mizutani and Mitsuoka, 1982). It could promote gastrointestinal (GI) inflammation (Jia et al., 2018) and change the composition of normal gut microbiota, by means of which bile acid becomes a risk factor during tumor formation (Jia et al., 2018). Administration of Konjac glucomannan was found to increase the fecal output of total bile acids in rats (Ikegami et al., 1984; Wu and Chen, 2011b). What's more, Konjac glucomannan significantly decreases secondary bile acid level in human subjects (Wu et al., 2011a). Therefore, the reduction of bile acids would be another functional aspect of glucomannan supplementation.

On the other hand, the probiotics, mainly *lactobacilli* and *bifidobacteria*, are indicated for their anti-colorectal cancer function through binding to the mutagens, inhibiting the activity of β -glucuronidase, and reducing the level of carcinogenic

secondary bile acid (Wu et al., 2011a; Chong, 2013). In various studies, glucomannan is shown to enhance the content of probiotics and its metabolic products, short-chain fatty acids (SCFAs), in feces (Mizutani and Mitsuoka, 1982; Ikegami et al., 1984; Yeh et al., 2007; Wu and Chen, 2011b; Wu et al., 2011a; Chong, 2013; Wu et al., 2014; Quezada et al., 2017). SCFAs promote the functionality of normal intestinal epithelium and the gut integrity (Chong, 2013); also, they stimulate cell differentiation program and induce apoptosis of transformed cells (Kumano et al., 1985; Yeh et al., 2007; Chong, 2013). Taken together, these results suggest that glucomannan could indirectly promote an environment that is unfavorable for cancer development, at least in the gastroenterological system (**Figure 2**).

Glucomannan Serves as a Targeted Carrier for Anti-Cancer Drug Delivery

A specific drug delivery system is necessary to enhance the efficacy of existing chemotherapeutic drugs and to reduce



their side effect. Glucomannan exhibits very low toxicity, modifiability, and the absence of immunogenicity (Zhan et al., 2014). These outstanding characteristics make it an ideal drug carrier.

Amphiphilic aliphatic amines grafted Konjac glucomannan micelle (Konjac glucomannan-g-AHs), a modified Konjac glucomannan, is added with a pH-sensitive Schiff's base. With improved stability, solubility, and cytotoxicity, the Konjacglucomannan-based micelles are able to protect the curcumin from degradation and deliver it into the tumor site through endocytosis-mediated transmembrane transport (Luan et al., 2017). Furthermore, glucomannan synergizes with other bioactive compounds to exert anti-tumor function. Combined with teniposide, they display enhanced cytotoxic and cytostatic effect on mouse leukemia cells (Miadoková et al., 2006) (Figure 3). In addition, the Konjac glucomannan/sodium alginate/graphene oxide (Konjac glucomannan/SA/GO) complex could effectively control the release of anti-cancer drug 5-FU in a local tumor site, thus reducing the systematic side effect (Wang et al., 2014; Yuan et al., 2019). Liu et al. designed a novel glucomannan-containing, aromatic azo agent bis(methacryloyl-amino)-azobenzene crosslinked hydrogel, which can release peptide drugs at the colon site in a pH-dependent or enzymatic degradation mode (Liu et al., 2004). Chitosan (CS)-coated microsphere, generated on the basis of oxidized Konjac glucomannan, makes another promising intestinespecific drug delivery approach for the treatment of bowel diseases, according to Shi's study (Shi et al., 2017). In brief, glucomannan is compatible with many other bioactive compounds and the conjugated macromolecular complexes could be easily taken by tumor cells, thus improving the therapeutic efficacy of existing anticancer chemicals.

The Effect of Glucomannan on a Tumor Immune Microenvironment

Progression of tumor into malignant stage is closely associated with a compromised immune surveillance function, wherein tumor-associated macrophages (TAMs) are predominantly involved (Zhan et al., 2014). Actually, glucomannan from Porang was demonstrated to activate macrophages through Toll-like receptor 4 (TLR4), increasing the production of interleukin-6 (IL-6) and tumor necrosis factor α (TNF α) and promoting its phagocytic capability in primary macrophages (Gurusmatika et al., 2017) (Figure 3). On the other hand, ubiquitous surface expression of glucose and mannose receptors makes TAMs possible targets for polysaccharides with anti-tumor activity (Chorvatovicova et al., 1999; Dong et al., 2009; Zhan et al., 2014). Although no specific study indicates that the glucomannan binding to the sugar receptor has direct influence on macrophage, studies emphasize the intermediary role of glucomannan as a TAM-targeting molecule. Glucomannan isolated from B. striata (BSP) shows nucleic acid binding affinity after modification by N,N'-carbonyl di-imidazole (CDI)/ethylene diamine (Zhan et al., 2014). The anti-sense nucleotides such as oligo-deoxynucleotide and small interference RNA (siRNA) could be delivered by the modified glucomannan via the recognition of macrophage surface receptors, which mediates the manipulation of TAMrelated gene expression level (Dong et al., 2009). A study shows



that glucomannan helps to selectively deliver alendronate to tumor-resident macrophages, thus leading to the apoptosis and depletion of TAMs (Zhan et al., 2014) (**Figure 3**). Also, the bisphosphonate glucomannan conjugate, consisting of polyethylene glycol (PEG), poly(lactic-co-glycolic acid) (PLGA), and a peptide that can be cleaved by matrix metalloproteases (MMPs), could be efficiently released to tumor sites and reduce the viability of TAMs.

Moreover, glucomannan was demonstrated to enhance the immune function both in vivo and in vitro (Zhan et al., 2014). Interleukin-10 (IL-10) is known as an immunosuppressive cytokine that is enriched in tumors to counteract the killing activity of cytotoxic T lymphocytes (CTLs) (Salazar-Onfray, 1999). Glucomannan could re-energize the immune system to attack the tumor cells by decreasing IL-10 level and promoting the production of IFN-y in tumor sites (Suzuki, 1983; Zhan et al., 2014). Similarly, the acidolysis-oxidized Konjac glucomannan up-regulates the expression of cytokines like TNFa, interleukin-1 β (IL-1 β), and IL-6, which collectively bolster the anti-tumor immune response (Vazquez-Velasco et al., 2015; Chen et al., 2017b) (Figure 3). However, it is important to note that glucomannan has also been suggested to possess an anti-inflammatory effect (Onishi et al., 2007; Wu et al., 2011a). Onishi N et al. indicated that pulverized konjac glucomannan suppressed the skin inflammatory immune response in NC/Nga mice evidenced by decrease of substance P, IL-10, IL-4, and TNF α (Onishi et al., 2007). Thus, whether the immune regulatory function of glucomannan is bidirectional or the immune motivating role is specific in local tumor environment remains to be determined by future studies.

DISCUSSION

As a straight-chain polymer with few branches, glucomannan was first appreciated for its role in gastroenterological disorders and metabolic diseases. Much similar to other polysaccharide extracts from traditional Chinese herbs, recently, the potential of glucomannan involved in anti-cancer therapy is being revealed. Accumulating evidence suggests that glucomannan exhibits broad but specific anti-tumor effect, when distinct cancer types are concerned (**Table 1**).

Mechanistically, glucomannan has direct impact on tumor cell survival and metastasis by blocking the PI3K/AKT signaling pathway (Frisch and Screaton, 2001; Guanen et al., 2018). After glucomannan treatment, cells show increased apoptosis along with decreased proliferation capacity. Meanwhile, the expression of chemokine receptors (CCR7 and CXCR4) is reduced, which

TABLE 1 | The anti-tumor mechanism of glucomannan in different cancer types.

Source of GM	Cancer type	Model		Mechanism
	Breast cancer (Wu et al., 2018; Wu et al.,	In vitro	MDA-MB-231	Induce cell cycle arrest
	2019)		MDA-MB-231BO	Inhibit migration and invasion
		In vivo	Mice	Regulation of the chromosomal and centrosomal instability
	Hepatoma (Ansil et al., 2013; Ansil et al.,	In vitro	PLC/PRF/5	Anti-proliferation
	2014c; Ansil et al., 2014b; Sawai et al., 2018)		HepG2	Increase apoptosis
		In vivo	Rat	Anti-oxidative stress
				Decrease cell viability
				Promote the production of propionate
		In vitro	SGC-7901	Increase apoptosis
Amorphophallus	Gastric cancer (Chen et al., 2017a)		AGS	Induce cell cycle arrest
konjac		In vivo	Human	Promote autophagy
	Colon carcinoma (Wu and Chen, 2011b; Ansil	In vitro	HCT-15	Induce apoptosis
	et al., 2013; Ansil et al., 2013; Ansil et al.,			Anti-proliferation
	2014a; Wu et al., 2014)	In vivo	Rat	Anti-oxidative stress
				Reduce β-glucuronidase and mucinase activities
				Promote the growth of bifidobacteria and lactobacilli and
				production of SCFAs
	Lung cancer	In vivo	Mice	-
Candida utilis	Lung cancer (Kumano et al., 1985)	In vivo	Mice	Inhibit migration and invasion
	Leukemia (Miadoková et al., 2006)	In vitro	P388D1	Anti-oxidative stress
Lentinus edodes	Ehrlich ascites tumor	In vivo	Mice	Induce expression of interferons
	Sarcoma S180-bearing (Fujii et al., 1978)			
Bletilla striata	Sarcoma S180-bearing (Zhan et al., 2014)	In vivo	Mice	Targeted depletion of TAMs
Aloe vera	Colon carcinoma (Im et al., 2016; Quezada	In vitro	HT29	Promote the growth of bifidobacteria and lactobacilli and
	et al., 2017)			production of SCFAs
		In vivo	Mice	Induce cell cycle arrest
	Leukemia (Sampedro et al., 2004)	In vitro	C1498	Anti-proliferation
	Ehrlich ascites tumor	In vivo	Mice	Activate macrophages
	Sarcoma 180-bearing (Im et al., 2005;			
	Liu et al., 2006)			

SCFAs, short-chain fatty acids; TAMs, tumor-associated macrophages.

Anti-Tumor Effect of Glucomannan

abrogates the migratory ability of the tumor cells (Wu et al., 2018). Uncontrolled cellular stress is pivotal in the early stage of tumorigenesis. Notably, hyperactive tumor metabolism leads to elevated ROS level and subsequent oxidative stress, which would cause genotoxicity and accumulation of mutations required for tumor development (Wu et al., 2014). Autophagy is a stress response characterized by degradation of self-components and the formation of autophagosomes. During early-stage tumor development, autophagy exerts a tumor-suppressive function through the clearance of stress-induced organelle damage. Glucomannan displays a protective effect by promoting the expression of genes associated with ROS scavenging and autophagy induction (Chen et al., 2017a). Though the underlying mechanism remains obscure, it is plausible to reason that the PI3K/AKT signaling pathway may also be critically involved in such stress responses. Studies show that inhibition of the PI3K/ AKT pathway limits the glycolytic process and deprives tumor cells of energy supply (Chen et al., 2016; Cretella et al., 2018), which could explain the reduction of ROS content (Robey and Hay, 2009; Zhao et al., 2017) and the elevation of cell autophagy level (Jin and White, 2007; Cheong, 2015; Liu et al., 2017). Altogether, glucomannan ameliorates cellular stress and prevents the tumorigenesis process.

Other than the direct interference of tumor cell biology, glucomannan also functions in an indirect manner. Glucomannan administration reduces carcinogen production in gut lumen and promotes probiotics and SCFA generation, which suppresses GI inflammation-related carcinogenesis (Wu et al., 2011a). On the other hand, due to its excellent bio-compatibility, glucomannan can work as a targeted delivery platform to enhance the efficiency of traditional drugs like curcumin (Luan et al., 2017), teniposide (Miadoková et al., 2006), and 5-FU (Wang et al., 2014). Anergic immune microenvironment is an important culprit in the process of tumor growth and evasion; thus, current therapies like checkpoint inhibitors are designed to motivate and re-energize the attacking ability of effector immune cells. Strikingly, glucomannan could alter the cytokine profile in the tumor vicinity and thus in support of CTLmediated cytotoxic effect. TAMs, tumor resident macrophages

REFERENCES

- Abramson, J. S., McGree, B., Noyes, S., Plummer, S., Wong, C., Chen, Y. B., et al. (2017). Anti-CD19 CAR T cells in CNS diffuse large-B-cell lymphoma. *N. Engl. J. Med.* 377 (8), 783–784. doi: 10.1056/NEJMc1704610
- Allemani, C., Matsuda, T., Di Carlo, V., Harewood, R., Matz, M., Niksic, M., et al. (2018). Global surveillance of trends in cancer survival 2000-14 (CONCORD-3): analysis of individual records for 37 513 025 patients diagnosed with one of 18 cancers from 322 population-based registries in 71 countries. *Lancet* 391 (10125), 1023–1075. doi: 10.1016/S0140-6736(17)33326-3
- Ansil, P. N., Prabha, S. P., Nitha, A., and Latha, M. S. (2013). Chemopreventive effect of *Amorphophallus campanulatus* (Roxb.) blume tuber against aberrant crypt foci and cell proliferation in 1, 2-dimethylhydrazine induced colon carcinogenesis. *Asian Pac. J. Cancer Prev.* 14 (9), 5331–5339. doi: 10.7314/ APJCP.2013.14.9.5331
- Ansil, P. N., Wills, P. J., Varun, R., and Latha, M. S. (2014a). Cytotoxic and apoptotic activities of *Amorphophallus campanulatus* (Roxb.) Bl. tuber extracts against

that contribute to the induction of a tolerant immune milieu, express receptors for various polysaccharide molecules. Glucomannan could increase macrophage phagocytic activity and cytokine secretion, such as TNF α and IL-6, *via* binding to TLR4 (Gurusmatika et al., 2017). It also facilitates the function of other compounds like siRNA (Dong et al., 2009) and alendronate (Zhan et al., 2014) to induce TAM apoptosis. All in all, further studies are needed to understand the details of how glucomannan impacts the immune system.

Cancer, the primary killer in modern society, is a complicated and systematic disease. The ultimate goal in cancer therapy is to completely eradicate tumor cells and reach the criteria of clinical cure without disease recurrence. It is a challenging task, considering the presence of cancer stem cells, intra-tumoral heterogeneity, and the formidable adaptability of tumor cells. While in pursuit of such purpose, it is also important to render patients live in "harmony" with their tumors, which means to maintain the high quality of life amid the effort to stop tumor from deterioration. According to a report published in JAMA Oncology in 2019, 1/3 of cancer patients would seek for help from alternative medicines (Sanford et al., 2019). Nature holds the key to cancer management, and ethnopharmacological herbs may be one of the answers that nature provides. Therefore, the identification, synthesization, and modification of additional bioactive anti-tumor compounds like glucomannan from natural ethnopharmacological herbs would greatly benefit the process of anti-tumor therapy and contribute to the improvement of human health.

AUTHOR CONTRIBUTIONS

J-YL, FS, and H-FZ proposed and wrote the manuscript. JY and CH collected and analyzed the information. HF supervised the conception and writing of the article.

FUNDING

The research was supported by the NSFC grant number 81774093.

human colon carcinoma cell line HCT-15. Saudi J. Biol. Sci. 21 (6), 524–531. doi: 10.1016/j.sjbs.2014.01.004

- Ansil, P. N., Wills, P. J., Varun, R., and Latha, M. S. (2014b). Cytotoxic and apoptotic activities of *Amorphophallus campanulatus* tuber extracts against human hepatoma cell line. *Res. Pharm. Sci.* 9 (4), 269–277. doi: 10.1016/j.sjbs.2014.01.004
- Ansil, P. N., Nitha, A., Prabha, S. P., and Latha, M. S. (2014c). Curative effect of *Amorphophallus campanulatus* (Roxb.) Blume. tuber on N-nitrosodiethylamineinduced hepatocellular carcinoma in rats. *J. Environ. Pathol. Toxicol. Oncol.* 33 (3), 205–218. doi: 10.1615/JEnvironPatholToxicolOncol.2014011320
- Bray, F., Ferlay, J., Soerjomataram, I., Siegel, R. L., Torre, L. A., and Jemal, A. (2018). Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J. Clin.* 68 (6), 394–424. doi: 10.3322/caac.21492
- Chen, L. L., Song, J. X., Lu, J. H., Yuan, Z. W., Liu, L. F., Durairajan, S. S., et al. (2014). Corynoxine, a natural autophagy enhancer, promotes the clearance of alpha-synuclein via Akt/mTOR pathway. J. Neuroimmune Pharmacol. 9 (3), 380–387. doi: 10.1007/s11481-014-9528-2

- Chen, X. S., Li, L. Y., Guan, Y. D., Yang, J. M., and Cheng, Y. (2016). Anticancer strategies based on the metabolic profile of tumor cells: therapeutic targeting of the Warburg effect. *Acta Pharmacol. Sin.* 37 (8), 1013–1019. doi: 10.1038/ aps.2016.47
- Chen, X., Yuan, L. Q., Li, L. J., Lv, Y., Chen, P. F., and Pan, L. (2017a). Suppression of gastric cancer by extract from the tuber of *Amorphophallus konjac via* induction of apoptosis and autophagy. *Oncol. Rep.* 38 (2), 1051–1058. doi: 10.3892/or.2017.5747
- Chen, M., Wang, S., Liang, X., Ma, D., He, L., and Liu, Y. (2017b). Effect of dietary acidolysis-oxidized Konjac glucomannan supplementation on serum immune parameters and intestinal immune-related gene expression of *Schizothorax prenanti. Int. J. Mol. Sci.* 18 (12), 2258. doi: 10.3390/ijms18122558
- Cheong, H. (2015). Integrating autophagy and metabolism in cancer. *Arch. Pharm. Res.* 38 (3), 358–371. doi: 10.1007/s12272-015-0562-2
- Chong, E. S. L. (2013). A potential role of probiotics in colorectal cancer prevention: review of possible mechanisms of action. J. Microbiol. Biotechnol. 30 (2), 351–374. doi: 10.1007/s11274-013-1499-6
- Chorvatovicova, D., Machova, E., Sandula, J., and Kogan, G. (1999). Protective effect of the yeast glucomannan against cyclophosphamide-induced mutagenicity. *Mutat. Res.* 444 (1), 117–122. doi: 10.1016/S1383-5718(99)00102-3
- CP C (2010). *Pharmacopoeia of the People's Republic of China*. 1st edition. Beijing: China Chemical Industry Press.
- Cretella, D., Ravelli, A., Fumarola, C., La Monica, S., Digiacomo, G., Cavazzoni, A., et al. (2018). The anti-tumor efficacy of CDK4/6 inhibition is enhanced by the combination with PI3K/AKT/mTOR inhibitors through impairment of glucose metabolism in TNBC cells. *J. Exp. Clin. Cancer Res.* 37 (1), 72. doi: 10.1186/s13046-018-0741-3
- Deng, S., Zhang, G., Kuai, J., Fan, P., Wang, X., Zhou, P., et al. (2018). Lentinan inhibits tumor angiogenesis via interferon gamma and in a T cell independent manner. J. Exp. Clin. Cancer Res. 37 (1), 260. doi: 10.1186/ s13046-018-0932-y
- Dong, L., Xia, S., Luo, Y., Diao, H., Zhang, J., Chen, J., et al. (2009). Targeting delivery oligonucleotide into macrophages by cationic polysaccharide from *Bletilla striata* successfully inhibited the expression of TNF-alpha. J. Control. Release 134 (3), 214–220. doi: 10.1016/j.jconrel.2008.11.013
- Frisch, S. M., and Screaton, R. A. (2001). Anoikis mechanisms. Curr. Opin. Cell Biol. 13 (5), 555–562. doi: 10.1016/S0955-0674(00)00251-9
- Fujii, T., Maeda, H., Suzuki, F., and Ishida, N. (1978). Isolation and characterization of a new antitumor polysaccharide, KS-2, extracted from culture mycelia of Lentinus edodes. J. Antibiot. (Tokyo) 31 (11), 1079–1090. doi: 10.7164/ antibiotics.31.1079
- Guanen, Q., Junjie, S., Baolin, W., Chaoyang, W., Yajuan, Y., Jing, L., et al. (2018). MiR-214 promotes cell meastasis and inhibites apoptosis of esophageal squamous cell carcinoma via PI3K/AKT/mTOR signaling pathway. *Biomed. Pharmacother.* 105, 350–361. doi: 10.1016/j.biopha.2018.05.149
- Gurusmatika, S., Nishi, K., Harmayani, E., Pranoto, Y., and Sugahara, T. (2017). Immunomodulatory activity of octenyl succinic anhydride modified Porang (*Amorphophallus oncophyllus*) glucomannan on mouse macrophage-like J774.1 cells and mouse primary peritoneal macrophages. *Molecules (Basel, Switzerland)* 22 (7), 1187. doi: 10.3390/molecules22071187
- Hassan, M., Watari, H., AbuAlmaaty, A., Ohba, Y., and Sakuragi, N. (2014). Apoptosis and molecular targeting therapy in cancer. *Biomed. Res. Int.* 2014, 150845. doi: 10.1155/2014/150845
- Ikegami, S., Harada, H., Tsuchihashi, N., Nagayama, S., Nishide, E., and Innami, S. (1984). Effect of indigestible polysaccharides on pancreatic exocrine secretion and biliary output. *J. Nutr. Sci. Vitaminol.* 30 (6), 515–523. doi: 10.3177/ jnsv.30.515
- Im, S. A., Oh, S. T., Song, S., Kim, M. R., Kim, D. S., Woo, S. S., et al. (2005). Identification of optimal molecular size of modified Aloe polysaccharides with maximum immunomodulatory activity. *Int. Immunopharmacol.* 5 (2), 271– 279. doi: 10.1016/j.intimp.2004.09.031
- Im, S. A., Kim, J. W., Kim, H. S., Park, C. S., Shin, E., Do, S. G., et al. (2016). Prevention of azoxymethane/dextran sodium sulfate-induced mouse colon carcinogenesis by processed Aloe vera gel. *Int. Immunopharmacol.* 40, 428– 435. doi: 10.1016/j.intimp.2016.09.022
- Jia, W., Xie, G., and Jia, W. (2018). Bile acid-microbiota crosstalk in gastrointestinal inflammation and carcinogenesis. *Nat. Rev. Gastroenterol. Hepatol.* 15 (2), 111– 128. doi: 10.1038/nrgastro.2017.119

- Jin, S., and White, E. (2007). Role of autophagy in cancer: management of metabolic stress. Autophagy 3 (1), 28–31. doi: 10.4161/auto.3269
- Kaiser, J. (2017). Personalized tumor vaccines keep cancer in check. *Science* 356 (6334), 122. doi: 10.1126/science.356.6334.122
- Kim, S. P., Kang, M. Y., Kim, J. H., Nam, S. H., and Friedman, M. (2011). Composition and mechanism of antitumor effects of *Hericium erinaceus* mushroom extracts in tumor-bearing mice. *J. Agric. Food Chem.* 59 (18), 9861– 9869. doi: 10.1021/jf201944n
- Klinder, A., Glei, M., and Pool-Zobel, B. (2008). Prebiotics and reduction of risk of carcinogenesis: Review of experimental and human data. Gibson G, Roberfroid M, editors. Boca Raton, FL: CRC Press. doi: 10.1201/9780849381829.ch15
- Kumano, N., Suzuki, S., Ishikawa, T., Koinumaru, S., and Konno, K. (1985). Antitumor effect of the yeast polysaccharide preparation in syngeneic mouse tumor models. *Tohoku J. Exp. Med.* 146 (1), 89–96. doi: 10.1620/tjem.146.89
- Liu, C., Leung, M. Y., Koon, J. C., Zhu, L. F., Hui, Y. Z., Yu, B., et al. (2006) Macrophage activation by polysaccharide biological response modifier isolated from Aloe vera L. var. chinensis (Haw.) Berg. *Int. Immunopharmacol.* 6 (11), 1634–1641. doi: 10.1016/j.intimp.2006.04.013
- Liu, Z.-L., Hu, H., and Zhuo, R.-X. (2004). Konjac glucomannan–graft–acrylic acid hydrogels containing azo crosslinker for colon-specific delivery. J. Polym. Sci. A 42 (17), 4370–4378. doi: 10.1002/pola.20272
- Liu, G., Pei, F., Yang, F., Li, L., Amin, A. D., Liu, S., et al. (2017). Role of autophagy and apoptosis in non-small-cell lung cancer. *Int. J. Mol. Sci.* 18 (2), 367. doi: 10.3390/ijms18020367
- Liu, D., Li, L., Zhang, X. X., Wan, D. Y., Xi, B. X., Hu, Z., et al. (2019). Correction: SIX1 promotes tumor lymphangiogenesis by coordinating TGFbeta signals that increase expression of VEGF-C. *Cancer Res.* 79 (7), 1715. doi: 10.1158/0008-5472.CAN-19-0462
- Luan, J., Wu, K., Li, C., Liu, J., Ni, X., Xiao, M., et al. (2017). pH-sensitive drug delivery system based on hydrophobic modified konjac glucomannan. *Carbohydr. Polym.* 171, 9–17. doi: 10.1016/j.carbpol.2017.04.094
- Miadoková, E., Svidová, S., Vlčková, V., Dúhová, V., Naďová, S., Rauko, P., et al. (2006). Diverse biomodulatory effects of glucomannan from *Candida utilis. Toxicol. In Vitro* 20 (5), 649–657. doi: 10.1016/j.tiv.2005.12.001
- Miller, R. S., and Hoskins, L. C. (1981). Mucin degradation in human colon ecosystems. Fecal population densities of mucin-degrading bacteria estimated by a "most probable number" method. *Gastroenterology* 81 (4), 759–765. doi: 10.1016/0016-5085(81)90503-5
- Mizutani, T., and Mitsuoka, T. (1982). Effect of Konjac mannan on spontaneous liver tumorigenesis and fecal flora in C3H/He male mice. *Cancer Lett.* 17 (1), 27–32. doi: 10.1016/0304-3835(82)90104-5
- Nassour, J., Radford, R., Correia, A., Fuste, J. M., Schoell, B., Jauch, A., et al. (2019). Autophagic cell death restricts chromosomal instability during replicative crisis. *Nature* 565 (7741), 659–663. doi: 10.1038/s41586-019-0885-0
- Onishi, N., Kawamoto, S., Suzuki, H., Santo, H., Aki, T., Shigeta, S., et al. (2007). Dietary pulverized konjac glucomannan suppresses scratching behavior and skin inflammatory immune responses in NC/Nga mice. *Int. Arch. Allergy Immunol.* 144 (2), 95–104. doi: 10.1159/000103220
- Quezada, M. P., Salinas, C., Gotteland, M., and Cardemil, L. (2017). Acemannan and fructans from Aloe vera (*Aloe barbadensis* Miller) plants as novel prebiotics. *J. Agric. Food Chem.* 65 (46), 10029–10039. doi: 10.1021/acs.jafc.7b04100
- Robey, R. B., and Hay, N. (2009). Is Akt the "Warburg kinase"?—Akt-energy metabolism interactions and oncogenesis. Sem. Cancer Biol. 19 (1), 25–31. doi: 10.1016/j.semcancer.2008.11.010
- Sabharwal, S. S., and Schumacker, P. T. (2014). Mitochondrial ROS in cancer: initiators, amplifiers or an Achilles' heel? *Nat. Rev. Cancer* 14 (11), 709–721. doi: 10.1038/nrc3803
- Salazar-Onfray, F. (1999). Interleukin-10: a cytokine used by tumors to escape immunosurveillance. Med. Oncol. (Northwood, London, England) 16 (2), 86–94. doi: 10.1007/BF02785841
- Sampedro, M. C., Artola, R. L., Murature, M., Murature, D., Ditamo, Y., Roth, G. A., et al. (2004). Mannan from *Aloe saponaria* inhibits tumoral cell activation and proliferation. *Int. Immunopharmacol.* 4 (3), 411–418. doi: 10.1016/j.intimp.2003.12.016
- Sanford, N. N., Sher, D. J., Ahn, C., Aizer, A. A., and Mahal, B. A. (2019). Prevalence and nondisclosure of complementary and alternative medicine use in patients with cancer and cancer survivors in the United States. *JAMA Oncol.* 5(5), 7350–737. doi: 10.1001/jamaoncol.2019.0349

- Sawai, S., Mohktar, M. S., Safwani, W., and Ramasamy, T. S. (2018). Suppression of the viability and proliferation of HepG2 hepatocellular carcinoma cell line by Konjac Glucomannan. *Anticancer Agents Med. Chem.* 18 (9), 1258–1266. doi: 10.2174/1871520618666180307143229
- Shi, M., Bai, J., Zhao, L., Yu, X., Liang, J., Liu, Y., et al. (2017). Co-loading and intestine-specific delivery of multiple antioxidants in pH-responsive microspheres based on TEMPO-oxidized polysaccharides. *Carbohydr. Polym.* 157, 858–865. doi: 10.1016/j.carbpol.2016.10.057
- Suzuki, S. (1983). Studies on the interferon-inducing activity of the yeast polysaccharide with reference to its antitumor effect in mice. Sci. Rep. Res. Inst. Tohoku Univ. Med. 30 (1–4), 56–64.
- Vazquez-Velasco, M., Gonzalez-Torres, L., Lopez-Gasco, P., Bastida, S., Benedi, J., Gonzalez-Munoz, M. J., et al. (2015). Effects of glucomannan/spirulina-surimi on liver oxidation and inflammation in Zucker rats fed atherogenic diets. *J. Physiol. Biochem.* 71 (4), 611–622. doi: 10.1007/s13105-015-0425-9
- Wang, J., Liu, C., Shuai, Y., Cui, X., and Nie, L. (2014). Controlled release of anticancer drug using graphene oxide as a drug-binding effector in konjac glucomannan/sodium alginate hydrogels. *Colloids Surf. B. Biointerfaces* 113, 223–229. doi: 10.1016/j.colsurfb.2013.09.009
- Wei, S. C., Levine, J. H., Cogdill, A. P., Zhao, Y., Anang, N. A. S., Andrews, M. C., et al. (2017). Distinct cellular mechanisms underlie anti-CTLA-4 and anti-PD-1 checkpoint blockade. *Cell* 170 (6), 1120–1133 e17. doi: 10.1016/j.cell.2017.07.024
- Wu, W.-T., and Chen, H.-L. (2011b). Effects of Konjac Glucomannan on putative risk factors for colon carcinogenesis in rats fed a high-fat diet. J. Agric. Food Chem. 59 (3), 989–994. doi: 10.1021/jf103532x
- Wu, W. T., Cheng, H. C., and Chen, H. L. (2011a). Ameliorative effects of konjac glucomannan on human faecal beta-glucuronidase activity, secondary bile acid levels and faecal water toxicity towards Caco-2 cells. *Br. J. Nutr.* 105 (4), 593–600. doi: 10.1017/S0007114510004009
- Wu, C., Chen, M., Zhang, Q., Yu, L., Zhu, J., and Gao, X. (2019). Genomic and GeneChip expression profiling reveals the inhibitory effects of Amorphophalli Rhizoma in TNBC cells. *J. Ethnopharmacol.* 235, 206–218. doi: 10.1016/j.jep.2019.02.004
- Wu, C., Qiu, S., Liu, P., Ge, Y., and Gao, X. (2018). Rhizoma Amorphophalli inhibits TNBC cell proliferation, migration, invasion and metastasis through the PI3K/Akt/mTOR pathway. J. Ethnopharmacol. 211, 89–100. doi: 10.1016/j. jep.2017.09.033
- Wu, W.-T., Yang, L.-C., and Chen, H.-L. (2014). Effects of konjac glucomannan, inulin and cellulose on acute colonic responses to genotoxic azoxymethane. *Food Chem.* 155, 304–310. doi: 10.1016/j.foodchem.2014.01.065

- Xiao, J. H., Zhang, Y., Liang, G. Y., Liu, R. M., Li, X. G., Zhang, L. T., et al. (2017). Synergistic antitumor efficacy of antibacterial helvolic acid from *Cordyceps taii* and cyclophosphamide in a tumor mouse model. *Exp. Biol. Med. (Maywood)* 242 (2), 214–222. doi: 10.1177/1535370216668051
- Yeh, S. L., Lin, M. S., and Chen, H. L. (2007). Inhibitory effects of a soluble dietary fiber from *Amorphophallus konjac* on cytotoxicity and DNA damage induced by fecal water in Caco-2 cells. *Planta Med.* 73 (13), 1384–1388. doi: 10.1055/s-2007-990228
- Yuan, Y., Xu, X., Gong, J., Mu, R., Li, Y., Wu, C., et al. (2019). Fabrication of chitosan-coated konjac glucomannan/sodium alginate/graphene oxide microspheres with enhanced colon-targeted delivery. *Int. J. Biol. Macromol.* 131, 209–217. doi: 10.1016/j.ijbiomac.2019.03.061
- Zhan, X., Jia, L., Niu, Y., Qi, H., Chen, X., Zhang, Q., et al. (2014). Targeted depletion of tumour-associated macrophages by an alendronate-glucomannan conjugate for cancer immunotherapy. *Biomaterials* 35 (38), 10046–10057. doi: 10.1016/j.biomaterials.2014.09.007
- Zhang, Y. (2017). Ganoderma lucidum (Reishi) suppresses proliferation and migration of breast cancer cells via inhibiting Wnt/beta-catenin signaling. Biochem. Biophys. Res. Commun. 488 (4), 679–684. doi: 10.1016/j. bbrc.2017.04.086
- Zhang, J., Li, H., Gao, D., Zhang, B., Zheng, M., Lun, M., et al. (2018). A prognosis and impact factor analysis of DC-CIK cell therapy for patients with hepatocellular carcinoma undergoing postoperative TACE. *Cancer Biol. Ther.* 19 (6), 475–483. doi: 10.1080/15384047.2018.1433501
- Zhao, Y., Hu, X., Liu, Y., Dong, S., Wen, Z., He, W., et al. (2017). ROS signaling under metabolic stress: cross-talk between AMPK and AKT pathway. *Mol. Cancer* 16 (1), 79. doi: 10.1186/s12943-017-0648-1

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2019 Li, Sun, Zhou, Yang, Huang and Fan. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.